



3D Nucleome Program

January, 2014
Council of Councils

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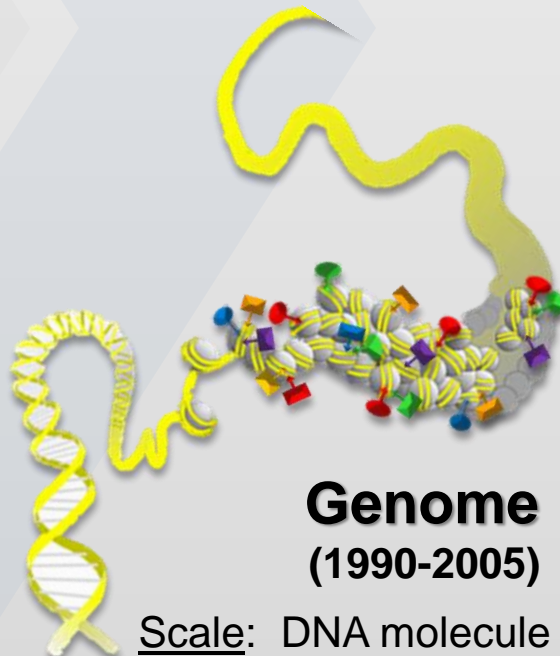
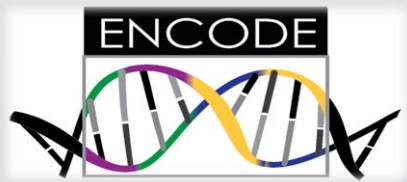
Finishing the Job: Understanding Genome Organization



Genome
(1990-2005)

Scale: DNA molecule &
sequence

Finishing the Job: Understanding Genome Organization



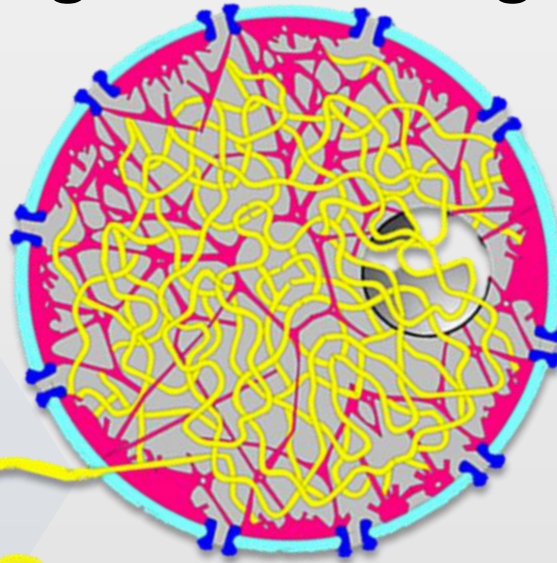
Genome
(1990-2005)

Scale: DNA molecule &
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Epigenome
(2005-2015)

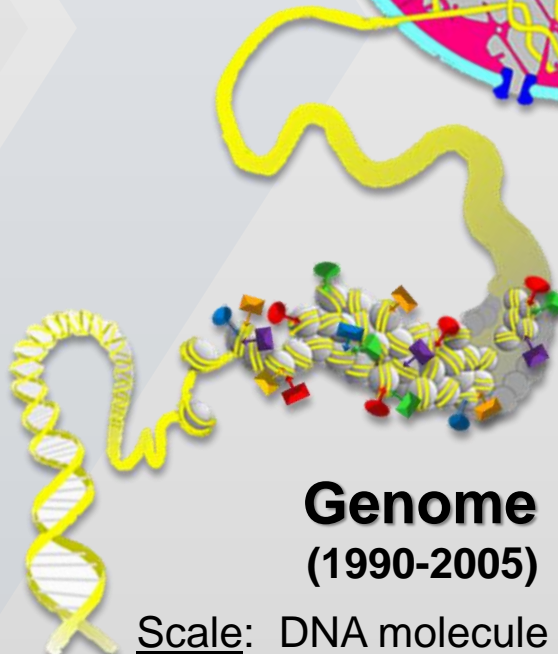
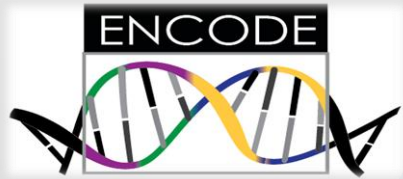
Scale: nucleosome &
epigenetic marks

Finishing the Job: Understanding Genome Organization



3D Nucleome (2015-2022?)

Scale: cell nucleus &
chromosome domains



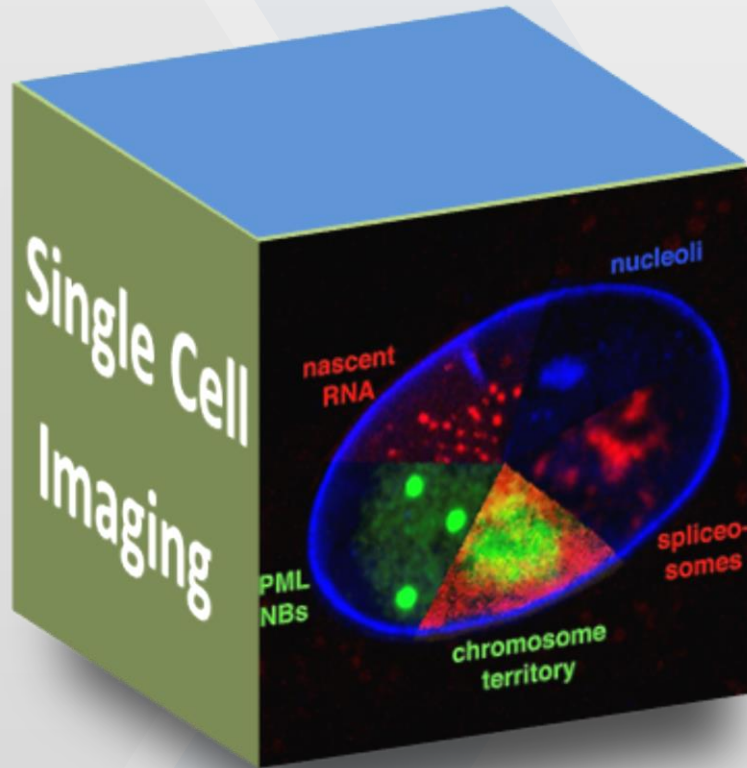
Genome (1990-2005)

Scale: DNA molecule &
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Epigenome (2005-2015)

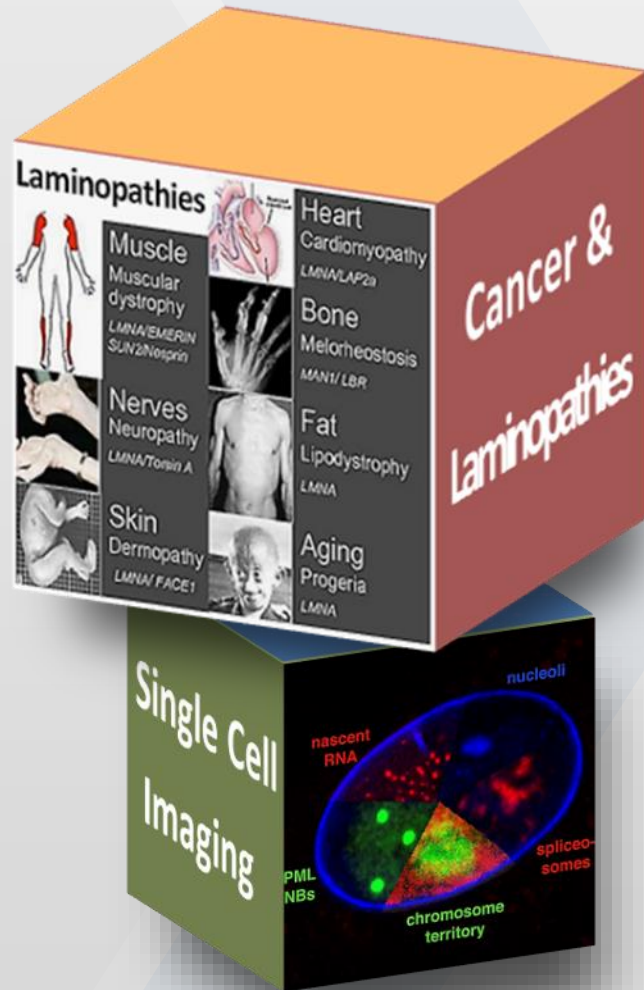
Scale: nucleosome &
epigenetic marks

Mounting Evidence: The Third Dimension Matters



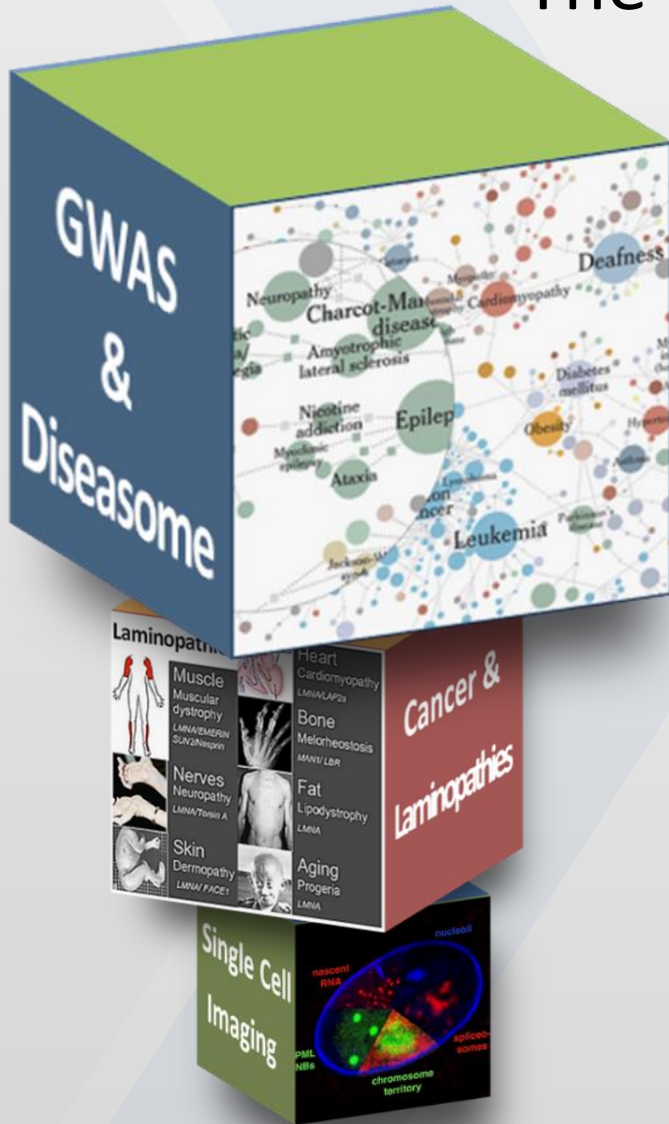
- The spatial distribution of the genome is not random;
- Chromatin is organized in chromosomal neighborhoods and associated with nuclear structures of unknown function;
- This organization is dynamic in time and space.

Mounting Evidence: The Third Dimension Matters



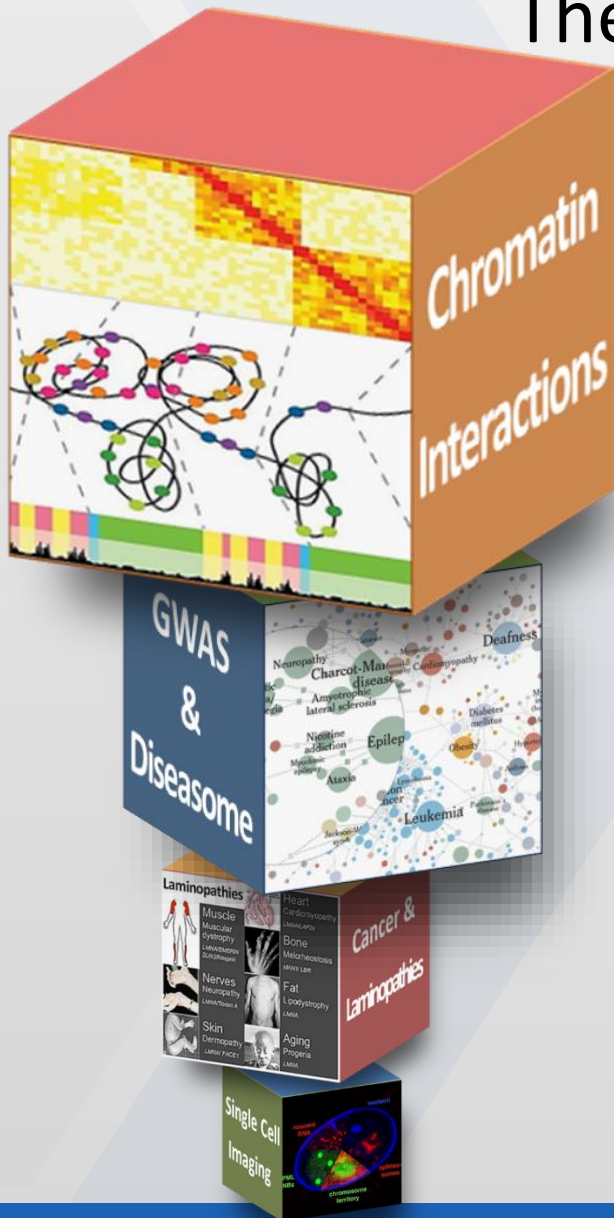
- Alterations of genome organization are associated with laminopathies, cancer and premature aging syndromes;
- Features are often cell- or tissue-specific, and include:
 - loss of genome integrity;
 - global changes in epigenetic marks;
 - widespread modifications of gene expression programs.

Mounting Evidence: The Third Dimension Matters



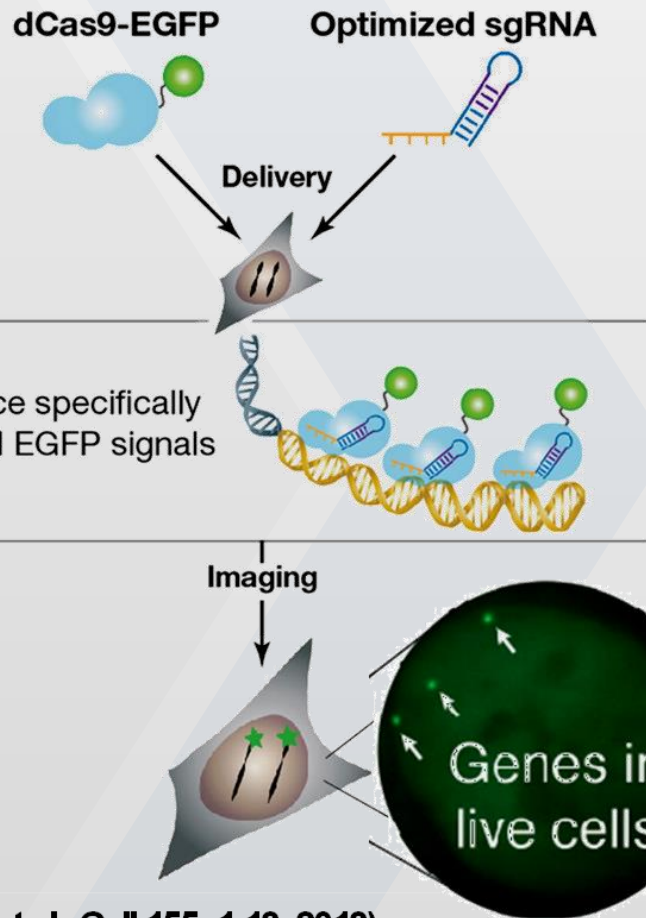
- 93% of disease-associated genetic variants reside in noncoding regulatory sequences with unknown targets;
- Mapping physical interactions between variants and promoters will help identify disease-associated target genes;
- Disease variants may belong to disease-specific modules within the nucleus.

Mounting Evidence: The Third Dimension Matters



- **Chromosome Conformation Capture (3C)** allows mapping of long-range looping interactions between genes and regulatory elements.
- First-generation maps suggest that the genome is organized in physically defined **Transcription Associated Domains (TADs)** and “transcription factories”.

A CRISPR Picture Of The Nuclear Genome



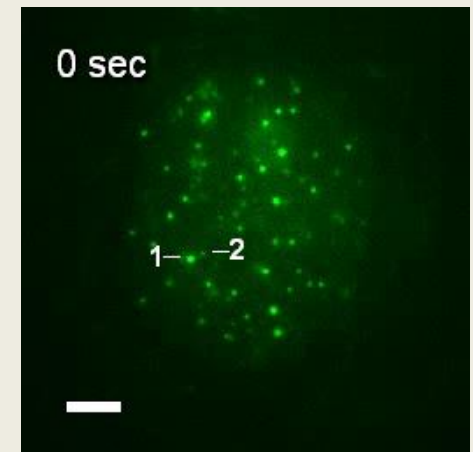
(Chen et al., Cell 155: 1-13, 2013)

The gene-editing **CRISPR/Cas** system can be used to:

- modify gene regions & regulatory elements in cells to test the organizational models inferred from the mapping of chromatin interactions;
- directly image genetic elements and follow them dynamically in live cells.

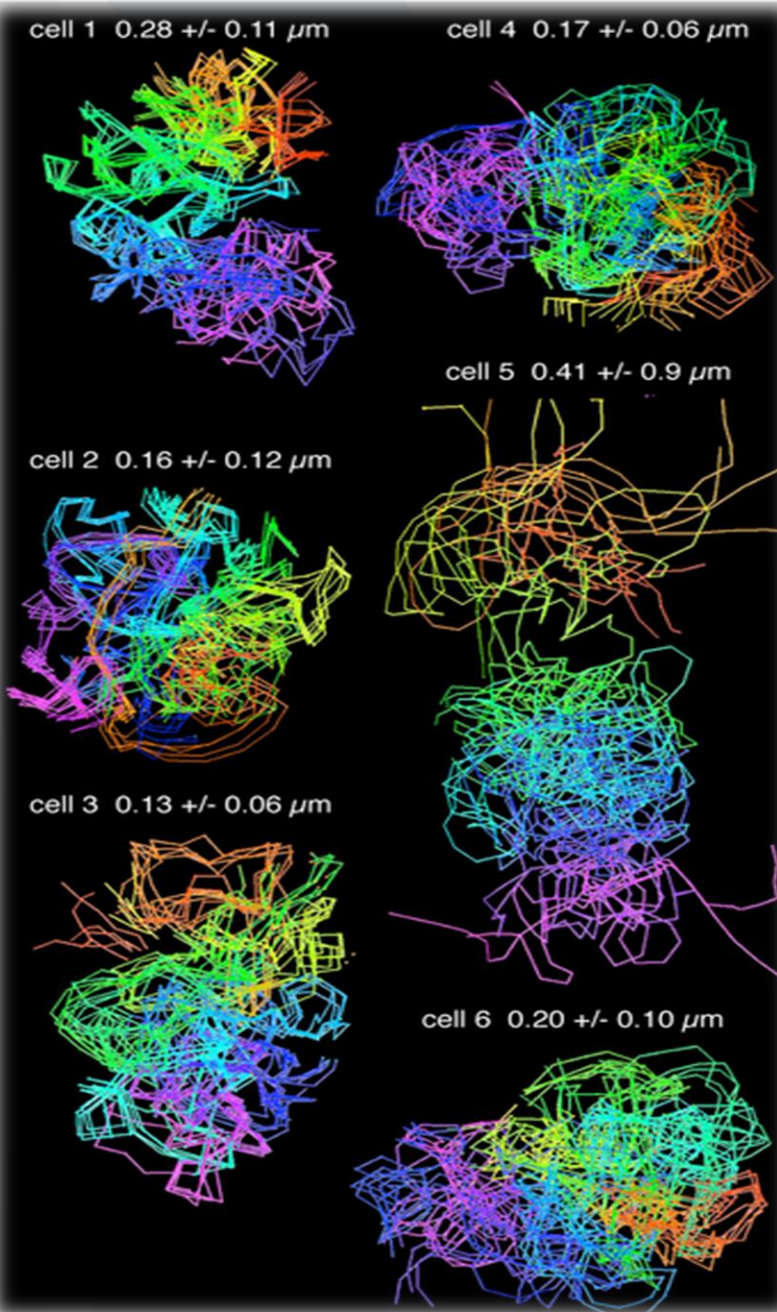
**Imaging
telomere
dynamics:**

(Lei Qi Lab, UCSF)



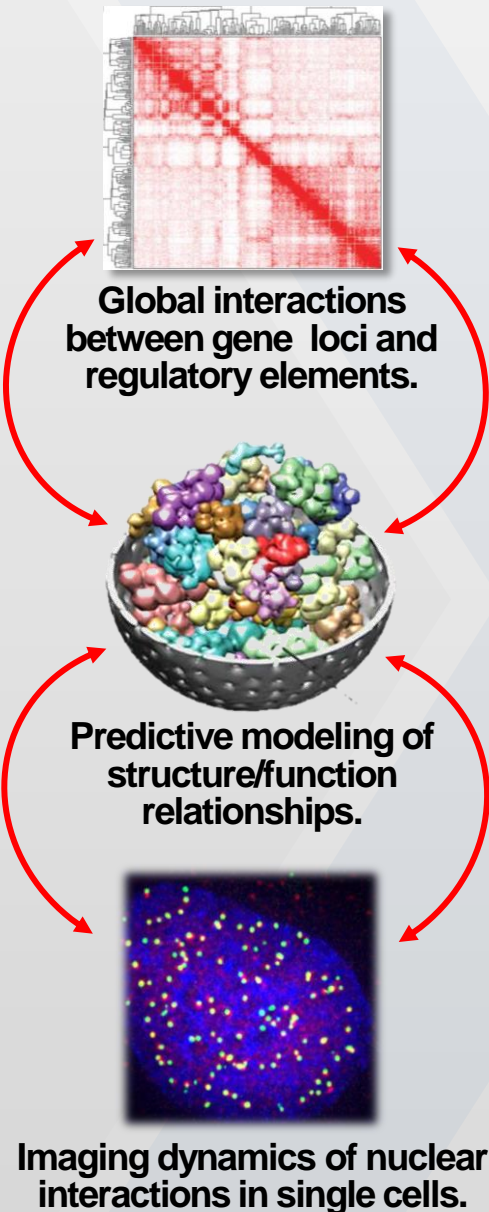
Single-cell Genome Conformation Capture (Hi-C)

- Single-cell Hi-C imaging can detect thousands of simultaneous chromatin contacts within a nucleus.
- Individual chromosomes maintain domain organization at the megabase scale, but chromosome structures vary from cell to cell at larger scales.



From Nagano T et al.,
Nature 502: 59–64,
03 October, 2013

Deliverables of the 3DN Program



- **Next generation tools** to explore the relationship between nuclear organization and regulation of gene expression programs in development and disease;
- **Reference maps** of the 3D architecture of the interphase nucleus for a variety of human cells and tissues;
- **Predictive models** of genome structure/function relationships;
- **Model validation** through controlled disruption of nuclear architecture and single-cell imaging.

Why We Need A Common Fund Program

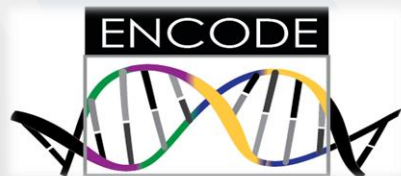


- Mapping the functional organization of the genome is **critical** to fully understand disease pathways and develop next generation diagnostics and therapeutics;
- 3DN tools and reference maps will **transform** many areas of biomedical research, but their development will require a **synergistic** effort;
- **Metrics and standards** need to be developed and adopted by a community of investigators, not just individuals.

3DN Initiatives, Timeline And Budgets

2015	2016	2017	2018	2019	2020	2021	2022
Mapping & Imaging Tools (\$5M/year) <ul style="list-style-type: none"> . High-resolution imaging tools, including single-cell. . New mapping technologies that do not rely on cross-linking. 				Structure/Function Relationships (\$5M/year) <ul style="list-style-type: none"> . Validate predictive models of structure/function relationships. . Study cell-cell variability and tissue-specificity. . Explore nuclear dynamics (4DN) in response to changes in the environment, signaling, cell division, cell differentiation. 			
Understand/Manipulate Nuclear Architecture (\$5M/year) <ul style="list-style-type: none"> . Study nuclear structures and their function. . Genome-editing technologies and engineered cell lines to allow controlled perturbation of 3DN. 				Next-Generation Tools (\$5M/year) <ul style="list-style-type: none"> . Single-cell imaging assays to follow dynamics of 100+ loci simultaneously. . In-vivo imaging of nuclear events in animal models (zebrafish, mouse). 			
Computational Tools (\$3M/year) <ul style="list-style-type: none"> . Increase resolution of mapping technologies. . Integrate physical interaction and imaging data. . Relate chromatin state maps to transcription programs. . Model 3DN structure / function relationships. 				Full-Scale Mapping: Reference Human 3DNs (\$10M/year) Data Coordinating Center: (\$1.5M/year) Mapping Consortium: (\$8.5M/year) <ul style="list-style-type: none"> . High-resolution whole-genome mapping of human 3DNs. . Reference maps of various cells types + in response to controlled environmental perturbations. . Public data repository and search tools for the community. 			
Pilot Mapping of Human 3DN (\$6M/year) Data Coordinating Center: (\$1.5M/year) . New DCC or supplement to ENCODE DCC. Mapping Consortium: (\$4.5M/year) <ul style="list-style-type: none"> . Pilot maps: low-resolution genome-wide AND high-resolution on 5% of human genome (depending on mapping technology). . Focus on reference cell lines (ENCODE). . Overlay ENCODE data on new 3DN physical maps. . Develop technologies, algorithms, metrics And standards as mapping progresses. . Formulate new hypotheses about the nuclear organization of the human genome. 				3DN & Disease Biology <ul style="list-style-type: none"> . Patient-derived iPSCs & role of 3DN in disease. . Revisit GWAS data in context of new 3DN maps + identify new disease/therapeutic targets; . Apply imaging technologies to IC-relevant cells and tissues. 			
Evaluation & decision on continuation.				Transition to IC-supported FOAs.			
\$19M	\$19M	\$19M	\$19M	\$20M	\$20M	\$20M	\$20M

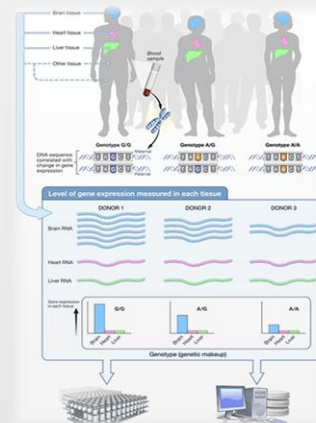
Building On U.S. And International Efforts



Encyclopedia Of DNA Elements (ENCODE)



Roadmap Epigenomics Program



Genotype – Tissue Expression (GTEx)



**Single Cell Analysis
Common Fund Program**



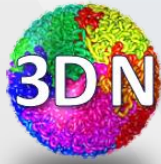
Haplotype Map of the human Genome (HapMap)



Library of Integrated Network-based Cellular Signatures (BD2K-LINCS)



The Cancer Genome Atlas (TCGA)



3D Nucleome

3DN Working Group

3DN Co-Chairs

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Phil Smith, **NIDDK**

3DN WG Members:

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Olivier Blondel, **NIDDK**

Lisa H. Chadwick, **NIEHS**

Richard Conroy, **NIBIB/Single Cell**

Max Guo, **NIA**

Sean Hanlon, **NCI**

Patricia Labosky, **DPCPSI**

Judy Mietz, **NCI**

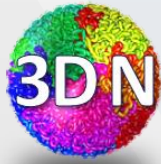
Mike Pazin, **NHGRI/ENCODE**

Matt Reilly, **NIAAA**

Robert Riddle, **NINDS**

Erica Rosemond, **NIMH**

John Satterlee, **NIDA/Epigenomics**



3D Nucleome

Expert Panel

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Mitch Guttman, Caltech

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